THE STRUCTURES OF SAIKOSAPONIN-E AND ACETYLSAIKOSAPONINS, MINOR COMPONENTS ISOLATED FROM <u>BUPLEURUM FALCATUM</u> L., DETERMINED BY C-13 NMR SPECTROSCOPY

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We report here a successful example of organic structure determination which requires several steps of chemical procedure and has readily been performed using ¹³C NMR spectroscopy without chemical degradations.

Triterpenoid glycosides, saikosaponin-a (5), $-b_1 - b_4$, -c, -d(2), and -f were already isolated from the methanolic extract of <u>Bupleurum falcatum</u> L. and their structures had been determined in this laboratory.¹⁻³ In connection with useful biological activities of this kind of saponins, we searched for minor components of <u>B. falcatum</u> and isolated seven saponins (1, 3, 4, 6, 7, 9, and 10) from the fractions with Rf values higher than those of the known saponins (see TABLE 1). Their 100-MHz ¹H NMR spectra in pyridine-d₅ revealed that five of them, 3, 4, 6, 7, and 9, have one OAc and six angular Me groups, and that 1 has seven angular Me groups and 10 only two angular Me groups (δ_H 0.60 and 0.74).

Recently, Tanaka and coworkers⁵ and we³ demonstrated that ¹³C NMR spectroscopy is a potential tool for elucidating structures of natural plant glycosides which are unstable against acid hydrolysis, provided



		1	3	4	6	2	2
mp		227-230°	219-225°	196-205°	217-224°	223-231°	209-217°
Rf ^a		0.25	0.58	0.40	0.33	0.22	0.31
Anal.		C ₄₂ H ₆₈ O ₁₂ · 2H ₂ O	C ₄₄ H ₇₀ O ₁₄ · 3H ₂ O	C ₄₄ H ₇₀ O ₁₄ ∙ H₂O	С ₄₄ Н ₇₀ О ₁₄ • Н ₂ О	С ₄₄ Н ₇₀ О ₁₄ · Н ₂ О	C ₄₅ H ₇₄ O ₁₅ ∙ 2H₂O
[α] _D (MeOH)		+40.8°	+47.5°	+43.5°	+44.3°	+47.6°	-6.4°
v ^{KBr} cm ⁻¹ max	(OAc)		1735	1735	1735	1715	1735
δ ^{C₅D₅N^b H}	OAc		1.95	1.96	1.96	2.12	1.95
	OMe						3.28
	H-6'	1.44	1.41	1.50	1.50	1.46	1.46

TABLE 1. Physical Properties of Saikosaponin-e (1) and Mono-Q-acetylsaikosaponins (3, 4, 6, 7, 9)

^a Solvent, EtOAc—EtOH—H₂O (9:1:0.5), double development. with a Varian HA-100 spectrometer at 100 MHz; $\delta_{H} \pm 0.01$. ^b ¹H NMR spectra were taken

that ¹³C data on their aglycones and sugar moieties are available, and presented some examples of glycosidation shifts, <u>i.e.</u>, chemical-shift changes from aglycone and sugar to saponin. The natural-abundance ¹H-noise-decoupled ¹³C FT NMR spectrum of <u>1</u>, a crystalline triterpenic glycoside named saikosaponin-e, readily revealed that it is a glucosyl



fucoside of saikogenin E, i.e., 13β , 28-epoxy- 16β -hydroxyolean-11-en- 3β - γ /Q- β -D-glucopyranosyl- $(1 \rightarrow 3)$ - β -D-fucopyranoside, upon comparison with the spectra of saikogenin E³ and saikosaponin-a (5).³ The ¹H NMR suggested that 10, $C_{35}H_{58}O_6$, mp 272-283°, $[\alpha]_D^{25}$ -30.0° (pyridine), is a glycoside of one of the sterol components already isolated from the plant.⁶ Thus, the ¹³C NMR of 10 was compared with those of authentic α -spinasterol and stigmasterol and revealed that 10 is α -spinasteryl β -D-glucopyranoside.⁷

On acetylation, the monoacetyl derivatives 3 and 4 afforded saikosaponin-d heptaacetate, 6 and 7 gave saikosaponin-a octaacetate, and 9 gave saikosaponin-b₄ nonaacetate. Since the ¹H NMR of 2-9 showed that the Me singlets of the new saponins, 3, 4, 6, and 9, appear at almost the same positions as those of the corresponding mother compounds, 2, 5, and 8, we inferred that their mono-Q-acetyl groups are situated at their sugar moieties. No other useful information was provided by their physical data. Moreover, it seemed laborious to determine the positions of their OAc groups using chemical techniques. However, the acetylation shift rule⁸ in ¹³C NMR spectroscopy can easily solve the problem, if the ¹³C signals of the mother compounds are fully assigned. We had assigned the signals of 2 and 5 in a

Carbon No.	<u>1</u> Δδ ^c	<u>ર</u> Δδ ₃₋₂	4 Δδ 4-2	<u>م</u> ک ⁸ ج	^Ζ Δδ _{ζ-Σ}	۶ ^{۵۵} ۲
1	39.0 (+0.2)	38.7 ^d (-0.1)	38.8 ^d (0.0)	39.0 (+0.1)	38.7 (-0.2)	40.2 (-0.1)
2	26.5 (-1.4)	25.9 (+0.1)	25.9 (+0.1)	25.8 (+0.2)	26.0 ^d (+0.4)	26.3 ^d (0.0)
3	88.9(+10.9)	82.5 (0.0)	82.5 (0.0)	82.4 (-0.1)	82.1 (-0.4)	82.8 (+0.1)
4	39.8 (+0.3)	43.9 (+0.1)	43.9 (+0.1)	43.7 (+0.2)	42.8 (-0.7)	43.8 (0.0)
5	55.9 (+0.6)	48.1 (0.0)	48.1 (0.0)	48.1 (+0.1)	48.9 (+0.9)	48.5 (0.0)
6	18.1 (-0.1)	17.9 (+0.1)	17.9 (+0.1)	17.9 (+0.1)	18.1 (+0.3)	18.6 (0.0)
7	32.2 (+0.3)	31.9 (+0.1)	31.9 (+0.1)	32.0 (+0.1)	32.1 (+0.2)	33.8 (-0.1)
8	42.5 (+0.3)	42.2 (+0.1)	42.2 (+0.1)	42.5 (+0.1)	42.5 (+0.1)	41.0 (0.0)
9	53.2 (+0.2)	53.3 (+0.1)	53.3 (+0.1)	53.3 (+0.1)	53.4 (+0.2)	53.5 ^e (-0.1)
10	36.7 (0.0)	36.7 (+0.1)	36.7 (+0.1)	36.7 (+0.1)	36.7 (+0.1)	38.6 (0.0)
11	132.0(-0.1)	132.0 (+0.1)	132.0 (+0.1)	132.0 (0.0)	131.8 (-0.2)	76.3 (0.0)
12	131.2(0.0)	132.0 (+0.1)	132.0 (+0.1)	131.1 (+0.1)	131.4 (+0.4)	122.7 (0.0)
13	84.1 (+0.1)	85.1 (0.0)	85.1 (0.0)	84.1 (+0.1)	84.1 (+0.1)	149.8 (0.0)
14	46.0 ^d (+0.4)	43.8 (+0.1)	43.8 (+0.1)	46.0 ^d (0.0)	$46.0^{e}(0.0)$	42.3 (0.0)
15	$36.2^{e}(-0.1)$	35.7 (0.0)	35.7(0.0)	36.4 (+0.2)	36.2 (0.0)	37.1 (-0.1)
16	64 4 (+0 4)	77.6 (+0.1)	77.6 (+0.1)	64.5 (+0.1)	64.4 (0.0)	74.3 (0.0)
17	47.0 ^d (0.0)	45.6 (+0.1)	45.6 (+0.1)	47.1 ^d (+0.2)	$47.1^{e}(+0.2)$	42.3 (0.0)
18	52 4 (+0 2)	$51 \land (+0 1)$	51 6 (+0.1)	52.4 (0.0)	52.4(0.0)	43.8 (0.0)
10	38 2 (+0 4)	38 9 ^d (+0 1)	38 9 ^d (+0 1)	38 2 (0 0)	38.2(0.0)	48 5 (0 0)
20	$31 \land (0.0)$	31 9 (+0 1)	31.9 (+0.1)	31.7 (+0.1)	31.7 (+0.1)	31.2 (0.0)
21	35.0 (+0.3)	37.1 (+0.1)	37.1 (+0.1)	35.0 (0.0)	35.0 (0.0)	35.2 (0.0)
22	25 7 ^e (00)	31 3 (+0 1)	31 3 (+0 1)	25 8 (+0 2)	25 8 ^d (+0 2)	30.0 (0.0)
23	28 1 (-0.3)	65 1 (-0 1)	65 2 (0 0)	65 0 (-0 1)	66 4 (+1 3)	65 6 (+0.1)
24	163(+0.4)	12 9 (+0 1)		12 8 (+0 1)	12.6(-0.1)	13 4 (-0 1)
25			18.8 (+0.1)	18 7 (+0 1)	18.4(-0.2)	$17.9^{f}(0.0)$
26	10.1 (-0.1) 19.9 (-0.1)	19 5 (+0 1)	19 5 (+0 1)	19.9 (+0.1)	20.0 (+0.2)	$18.8^{f}(0.0)$
20	20.9(0.0)	18 2 (+0.1)	18 2 (+0 1)	20.9(+0.1)	20.8 (0.0)	$26.5^{d}(0.0)$
28		77 9 (+0 1)	77.8 (0.0)	72 9 (0 0)	73 1 (+0 2)	70 2 (0 0)
20	33.7(0.0)	33.7(0.0)	33 7 (0 0)	33.7(0.0)	33 7 (0 0)	33 2 (0 0)
20	22 Q (±0,1)	33.7 (0.0)	24 6 40 1)	$23 \circ (0.0)$	23 9 (0 0)	
50	20.7 (10.1)	24.0 (10.1)	24.0 (0.1)	23.7 (0.0)	20.7 (0.0)	23.0 (0.0)
1'	106.3 (—)	105.6 (+0.2)	105.6 (+0.2)	105.6 (+0.4)	106.0 (<u>+0.8</u>)	105.6 (+0.1)
2'	71.6 (—)	71.8 (+0.1)	71.6 (-0.1)	71.5 (-0.2)	71.6 (-0.1)	71.6 (-0.2)
3'	85.1 (—)	85.2 (+0.1)	85.4 (+0.3)	85.4 (+0.4)	85.3 (+0.3)	85.4 (+0.1)
4'	71.9 (—)	72.1 (+0.2)	71.7 (-0.2)	71.7 (0.0)	72.1 (+0.4)	71.7 (-0.3)
5'	70.8 (—)	71.0 (+0.1)	71.0 (+0.1)	71.0 (+0.2)	71.0 (+0.2)	71.0 (0.0)
6'	17.0 (—)	17.0 (+0.1)	17.1 (+0 <i>.</i> 2)	17.1 (+0.2)	17.0 (+0.1)	17.0 (0.0)
1.1	105.9(-)	105 0 (+0 1)	105 6 (-0.2)	105 6 (+0 1)	106 0 (+0 5)	105 6 (-0.3)
2"	75.5 ()	73.7(-1.8)	$75 \land (-0.1)$	75 4 (+0.1)	75 7 (+0.4)	75 3 (-0.3)
3"	73.3(-)	79.5 (+1.4)	78.1 (0.0)	78.1 (+0.2)	78.3 (+0.4)	78.1 (-0.2)
۵. ۲۳	71.9(-)	70.0(-1.9)	72.0 (+0.1)	72.0 (+0.3)	72.1 (+0.4)	72.0 (0.0)
5"	78.1(-)	78.3 (+0.2)	75.4 (-2.7)	75.4 (-2.5)	78.3 (+0.4)	75.3 (-3.0)
6"	62.9 ()	62.6 (-0.4)	64.7 (+1.7)	64.6 (+1.8)	63.1 (+0.3)	64.6 (+1.6)
Maco	····· ()	21.0(-)	20.6 ()	20.6 ()	20.8 ()	20.6 (
MeCO		170.8()	170.6(-)	170.4()	170.5()	170.7(-)
MeŌ						51.8 ^e (0.0)
						51.5 (0.0)

TABLE 2. Carbon-13 Chemical Shifts $\delta_{C} (\pm 0.1)^{a}$ of Saikosaponin-e (1) and Mono-Qacetylsaikosaponins (3, 4, 6, 7, 9), and Acetylation Shifts $\Delta \delta$ (in parentheses, ppm)^b

TABLE 2 (continued)

^a ¹³C NMR spectra were recorded on a Varian NV-14 FT NMR spectrometer at 15.087 MHz in pyridine-d₅ with TMS as an internal reference (δ_{C} 0) in 8-mm spinning tubes at 100°; concentrations were about 0.15 mmol/cm³. FT measurement conditions were as follows: spectral width, 3923 Hz; pulse flipping angle, <u>ca</u>. 36°; acquisition time, 0.6 sec; number of data points, 4820. ^b See text; plus sign denotes a downfield shift. ^c Glycosidation shifts from saikogenin E (see ref 3). ^{d-f} Assignments may be interchanged in each vertical column.

structural study of other saikosaponins.³ The signal assignment for 8 was straightforward.

Thus, ¹³C spectra of the mono-Q-acetylsaikosaponins were examined in pyridine-d₅ at 100° and compared with those of their mother compounds. TABLE 2 shows that the signals of α -carbons carrying an acetylated OH group shifted downfield by +1.3~+1.8 ppm and the signals of β -carbons moved upfield by -0.7~-3.0 ppm. Therefore, the positions of the OAc groups were readily pointed out, <u>i.e.</u>, compounds 3, 4, 6, 7, and 9 are 3"-Q-acetyl- and 6"-Q-acetylsaikosaponins-d, 6"-Q-acetyl- and 23-Q-acetylsaikosaponins-a, and 6"-Q-acetylsaikosaponin-b₄ (this appears to be an artifact),² respectively.⁹

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REFERENCES

- (1) T. Kubota and H. Hinoh, Tetrahedron Lett. 303 (1968).
- (2) A. Shimaoka, S. Seo and H. Minato, J.C.S. Perkin I 2043 (1975).
- (3) K. Tori, Y. Yoshimura, S. Seo, K. Sakurawi, Y. Tomita and H. Ishii, <u>Tetrahedron Lett.</u> 4163 (1976); K. Tori, S. Seo, Y. Yoshimura, M. Nakamura, Y. Tomita and H. Ishii, <u>Ibid.</u> 4167 (1976).
- (4) M. Yamamoto, A. Kumagai and Y. Yamamura, <u>Arzneim.-Forsch. (Drug Res.)</u> <u>25</u>, 1021 (1975); G. S. Rao, J. E. Sinsheimer and K. W. Cochran, <u>J. Pharm. Sci.</u> <u>63</u>, 471 (1974).
- (5) K. Yamasaki, H. Kohda, T. Kobayashi, R. Kasai and O. Tanaka, Tetrahedron Lett. 1005 (1976).
- (6) K. Takeda, K. Hamamoto and T. Kubota, Yakugaku Zasshi (J. Pharm. Soc. Japan) <u>73</u>, 272 (1953);
 K. Takeda and T. Kubota, Chem. Pharm. Bull. <u>6</u>, 536 (1958).
- (7) M. Ito, K. Nishimoto, S. Natori and N. Ikekawa, <u>Shoyakugaku Zasshi (Japan. J. Pharmacog.)</u> <u>18</u>, 16 (1964).
- (8) As a leading reference, see Y. Terui, K. Tori and N. Tsuji, <u>Tetrahedron Lett.</u> 621 (1976).
- (9) Compound 3 is identical with that isolated from the same plant by Tanaka, et al. 10
- (10) K. Yamasaki, R. Kasai, Y. Masaki, M. Okihara, O. Tanaka, H. Oshio, S. Takagi, M. Yamaki, K. Masuda, G. Nonaka, M. Tsuboi and I. Nishioka, <u>Tetrahedron Lett.</u> the following paper.